

## PREPARATION AND IMMUNOSUPPRESSIVE ACTIVITY OF 32-(O)-ACYLATED AND 32-(O)-THIOACYLATED ANALOGUES OF ASCOMYCIN

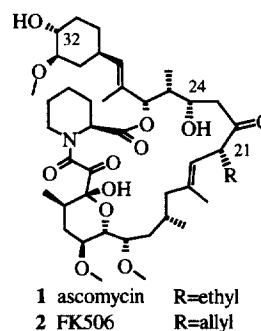
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**Abstract:** A series of 32-(O)-acylated and 32-(O)-thioacylated derivatives of the antibiotic ascomycin (**1**) have been synthesized. These readily accessible analogues exhibit potent immunosuppressive activity *in vitro*, as measured by an interleukin-2 reporter gene assay and the mixed lymphocyte reaction. Such molecules are expected to have a therapeutic potential in chronic inflammatory diseases of the airways such as asthma. © 1999 Elsevier Science Ltd. All rights reserved.

The antifungal antibiotic ascomycin (**1**) has been known for more than 30 years<sup>1</sup>. Both ascomycin and the better known allyl analogue, the immunosuppressant FK506 (**2**), are powerful inhibitors of antigen-stimulated T-cell activation and proliferation<sup>2</sup>. Similar to the chemically different cyclosporin A (CsA), ascomycin and its analogues when bound to the intracellular binding protein FKBP-12, block the signal transduction pathway in T-cells through inhibiting the Ca-dependent protein phosphatase calcineurin<sup>3</sup>. As a consequence, translocation of the transcription factor NF-AT from the cytoplasm to the nucleus is blocked, resulting in a failure to activate genes necessary for T-cell proliferation (e.g. interleukin-2). Both FK506 and CsA are successfully used in transplantation for prevention of organ graft rejection<sup>2,4</sup>. Therapeutic effects with CsA have also been observed in rheumatoid arthritis<sup>5</sup> and psoriasis<sup>6</sup>. Furthermore, recent preclinical data indicate that selective T-cell inhibitors such as CsA and ascomycin analogues could be of interest in treating asthma, a chronic inflammatory disease of the airways<sup>7</sup>. Indeed, oral CsA has been shown to be of clinical benefit in severe asthmatic patients<sup>8</sup>. These findings are in line with the current view that the T-cell plays a pivotal role in the pathology of asthma<sup>9</sup>. However, due to their inherent, mechanistically related side effects there might be a concern of using current immunosuppressants such as FK506 and CsA as an oral therapy for non-life threatening asthma<sup>10</sup>. Systemic exposure and hence the likelihood of untoward side effects could be reduced by giving such compounds locally by inhalation<sup>11</sup>. Furthermore, in contrast to FK506 and CsA, the ideal immunosuppressant should have no oral activity because current inhalation devices cannot prevent the majority of the inhaled drug (80-90%) from being deposited in the upper airways, and finally being swallowed. We therefore initiated a program with the objective of finding simple, potent immunosuppressive and anti-inflammatory ascomycin analogues which would be suited for inhalation therapy of respiratory diseases such as

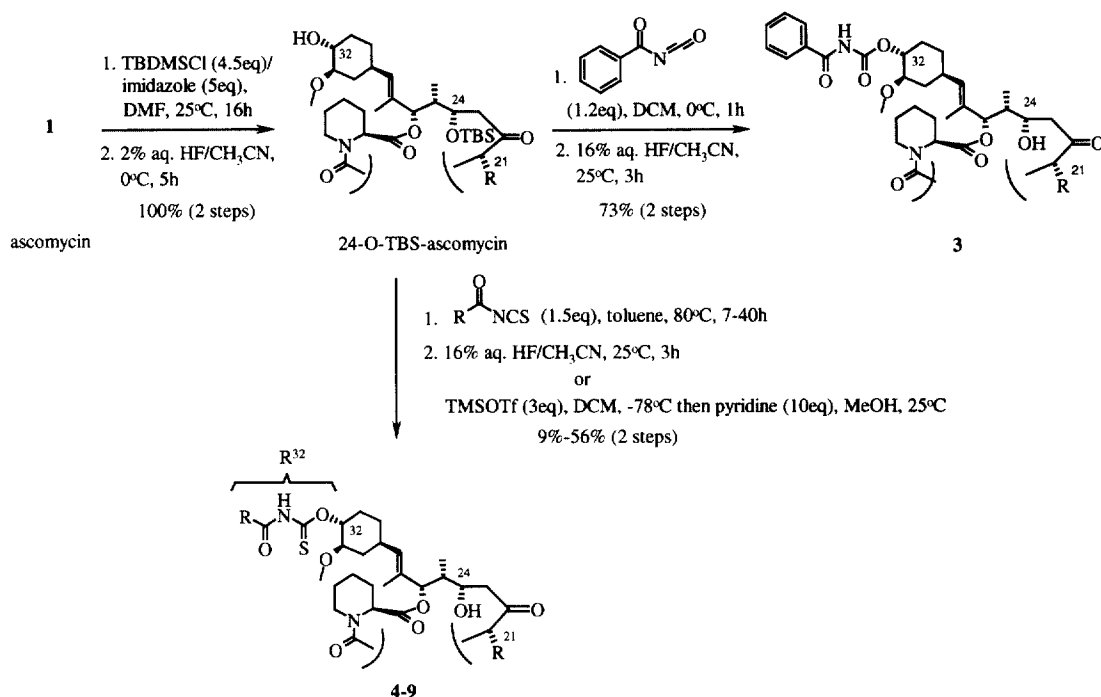


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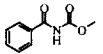
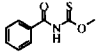
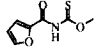
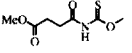
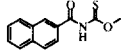
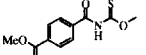
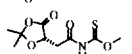
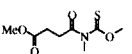
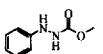
asthma. This paper reports the synthesis and the biological characterization *in vitro* of a series of easily accessible 32-(*O*)-acylated and 32-(*O*)-thioacylated analogues of ascomycin (**1**).

**Chemistry:** The synthesis of benzoyl carbamic acid 32-(*O*)-ascomycinyl ester **3** and of the acyl thiocarbamic acid 32-(*O*)-ascomycinyl esters **4–9** is illustrated in Scheme 1 and Table 1. Protection of the competing 24-hydroxyl group as the TBS-ether turned out to be advantageous and was accomplished in quantitative yield by a bis-silylation/ monodesilylation protocol. Reaction of the mono-protected ascomycin with acyl isothiocyanates (Table 2) in toluene or benzoyl isocyanate in dichloromethane, followed by deprotection using aqueous HF in acetonitrile afforded compounds **3–8** in moderate yields. A special deprotection protocol was applied for the synthesis of compound **9**. In order to prevent dioxolanone ring-opening under aqueous acidic conditions, the silyl ether was selectively cleaved with trimethylsilyl triflate in dichloromethane at  $-78^{\circ}\text{C}$ <sup>12</sup>. The synthesis of non-commercially available acyl isothiocyanates is illustrated in Table 2. They were made from the corresponding acid chlorides by reaction with sodium thiocyanate in ethyl acetate<sup>13</sup>. No purification step was necessary and, after removal of sodium chloride by filtration, the products were obtained in moderate to good yields and sufficiently pure for direct use.

Scheme 1



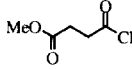
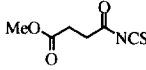
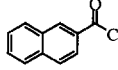
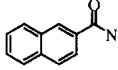
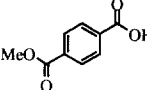
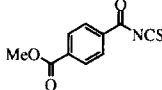
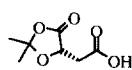
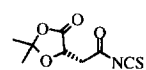


**Table 1:** Structures and *in vitro* immunosuppressive activities of compounds 1–11

compound	R32	R21	IL-2 RGA rIC <sub>50</sub> <sup>a</sup>	MLR rIC <sub>50</sub> <sup>a</sup>	FKBP-12 rIC <sub>50</sub> <sup>a</sup>
1 ascomycin	OH	ethyl	1.7	1.9	0.9
2 FK506	OH	allyl	1	1	1
3		ethyl	1.2	2.9	1.3
4		ethyl	3.4	11	6.3
5		ethyl	3.5	7.6	3.4
6		ethyl	4.6	19	1.6
7		ethyl	6.5	15	6.6
8		ethyl	9.0	8.4	7.3
9		ethyl	12	12	0.9
10		ethyl	6.9	36	2.7
11		ethyl	0.7	2.5	1.5

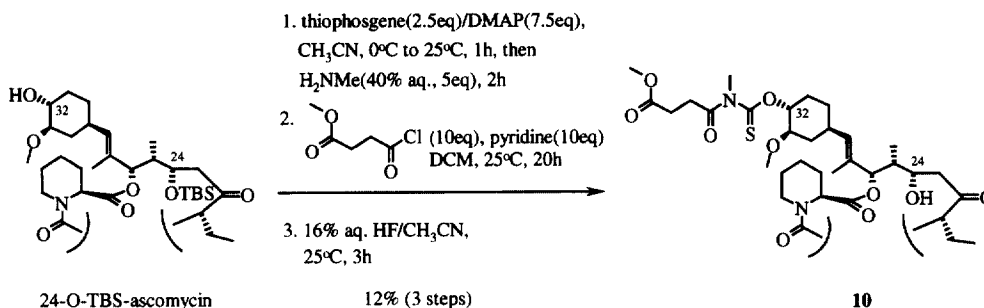
<sup>a</sup> Mean IC<sub>50</sub> [nM] values (n=2–8) of the test compounds are compared to that of FK506 and expressed as relative IC<sub>50</sub>: rIC<sub>50</sub>=IC<sub>50</sub> substance / IC<sub>50</sub> FK506. The absolute IC<sub>50</sub> values for FK506 in the IL-2 RGA, the MLR and the FKBP-12 binding assay are 0.2nM, 0.3nM and 1.2nM, respectively.

**Table 2:** General synthesis of acyl isothiocyanates

$\text{R}-\text{COOH} \xrightarrow[\text{toluene, 70}^\circ\text{C}]{\text{SOCl}_2} \text{R}-\text{COCl} \xrightarrow[\text{AcOEt, 0}^\circ\text{C}]{\text{NaSCN (1.2eq)}} \text{R}-\text{C(=O)NCS}$		
starting material	product	yield
		21% (2steps)
		95%
		95%
		73% (2steps)
		64% (2steps)

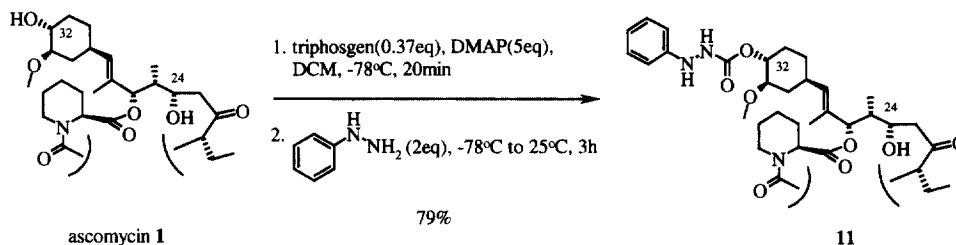
The synthesis of compound **10** is illustrated in Scheme 2. 24-(*O*)-TBS-ascomycin was first converted to the corresponding *N*-methyl-thiocarbamic acid 32-(*O*)-ascomycinyl ester by activation with thiophosgene/4-dimethylaminopyridine in acetonitrile and *in situ* reaction with aqueous methylamine. Acylation of the *N*-methyl-thiocarbamate intermediate with 3-chlorocarbonyl propionic acid methyl ester followed by the usual deprotection with aqueous HF led to product **10**.

Scheme 2



Finally, Scheme 3 illustrates the synthesis of compound **11**. Remarkably, no 24-hydroxyl protection was necessary in this case and selective 32-hydroxyl mono-acylation was achieved by activation of ascomycin (**1**) at -78°C with triphosgen/4-dimethylaminopyridine followed by *in situ* reaction with phenylhydrazine.

Scheme 3



**Results and Discussion:** Table 1 illustrates the *in vitro* immunosuppressive activity of **1–11** as measured in the interleukin-2 reporter gene assay (IL-2 RGA) and the mouse mixed lymphocyte reaction (MLR)<sup>14</sup>. Both assays are models for *in vitro* T-cell activation. In the case of the IL-2 RGA the activity of the IL-2 promoter in a

mitogen-stimulated human T-cell line is determined. The MLR is a model for T-cell activation by alloantigen. Since binding to FKBP-12 is a prerequisite for the immunosuppressive activity of ascomycin analogues the results for binding to FKBP-12 are also shown. All values are expressed as relative activities to FK506 (**2**) which is about twice as potent as the parent ascomycin (**1**)<sup>14</sup>. The benzoyl carbamic acid ester derivative **3** was equipotent to ascomycin (**1**) in both cellular assays the IL-2 RGA and the MLR. Consistent with this, analogue **3** and ascomycin (**1**) showed similar activity in the binding to FKBP-12. Replacement of the oxygen atom by sulfur in the R<sup>32</sup> linker led to a decrease of immunosuppressive activity as illustrated by direct comparison of compounds **3** and **4**. In general, increasing the size of the R<sup>32</sup> substituent in the acyl thiocarbamic acid ester series **4–10** gave less potent compounds although the loss of activity was only moderate ( $\leq 10$ ). This indicates that there is some degree of steric tolerance at position 32 of ascomycin which allows chemical modification of the compound. This has also been observed by others<sup>15</sup> and may be explained by the structure of the active drug/FKBP-12/calcineurin complex<sup>16</sup>. It is noteworthy that compound **10** was only about twice less potent than the non *N*-methylated analogue **6**. Thus, the NH bond of the carbamic acid ester linker seems not to be involved in any significant hydrogen-bond interaction with the target proteins. The most potent compound synthesized in this series was derivative **11**. Replacing the 32-hydroxy group of ascomycin (**1**) by the larger *N'*-phenylhydrazinecarboxylic acid ester moiety doubled the potency of the parent compound in the IL-2 RGA and retained the activity in the MLR. It is noteworthy, that the IC<sub>50</sub> values determined with human T-cells (IL-2 RGA) were comparable with the IC<sub>50</sub> values determined with mouse T-cells (MLR) differing only by a factor of 1 to 5. As reported previously<sup>17</sup>, and confirmed in this study, binding to FKBP-12 is a prerequisite for immunosuppressive activity of ascomycin analogues but not necessarily predictive for their immunosuppressive efficacy *in vitro*. For example, compound **9** was equipotent to compound **3** and parent ascomycin (**1**) with respect to binding to FKBP-12 but was about 10 times weaker in the IL-2 RGA and the MLR. A possible explanation for this finding is that the dioxolanone-substituent strengthens the macrolide/FKBP-12 interaction as indicated by the low rIC<sub>50</sub>, but weakens the affinity of the macrolide/FKBP-12 binary complex for its cellular target, the protein phosphatase calcineurin thereby reducing the immunosuppressive activity of **9**.

In summary, we have synthesized a series of easily accessible 32-(*O*)-acylated and 32-(*O*)-thioacylated ascomycin derivatives and evaluated their immunosuppressive properties *in vitro* in the IL-2 RGA, the MLR and the FKBP-12 binding assay. Modification of the 32-hydroxy group of ascomycin (**1**) led to the identification of analogues (e.g. **3** and **11**) whose immunosuppressive activity was equal or better than that of the parent natural product. Further SAR of 32-carbamic and thiocarbamic acid ascomycinyl ester analogues as well as their characterization in *in vivo* models of allergic asthma are ongoing and will be the subject of a future publication.

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In all assays the mean IC<sub>50</sub> [nM] value of the test compound (n=2-8) was compared to that of FK506 and expressed as relative IC<sub>50</sub>: rIC<sub>50</sub>=IC<sub>50</sub> substance / IC<sub>50</sub> FK506. The absolute IC<sub>50</sub> values for FK506 in the IL-2 RGA, the MLR and the MBA are 0.2nM, 0.3nM and 1.2nM, respectively.
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